

Determination of iron, copper, manganese, zinc, lead, and cadmium in mushroom samples from Tokat, Turkey

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Abstract

Contents of Fe, Cu, Mn, Zn, Pb, and Cd in 10 mushrooms species from Tokat, Turkey were determined by atomic absorption spectrometry. The results indicate that Fe, Cu, Mn, and Pb levels in the species *Fomes fomentarius* were the highest with means of 3904 ± 307 mg/kg, 54 ± 4 mg/kg, 64 ± 5 mg/kg, 2.7 ± 2.0 mg/kg, respectively. The highest level of Zn was 122 ± 11 mg/kg in the species *Polyporus frondosus*. Content of Cd in *Boletus appendiculatus* and *Fomes fomentarius* were the highest with a mean of 1.8 ± 0.2 mg/kg.

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1. Introduction

Mushrooms are important in the ecosystem because they are able to biodegrade the substrate and therefore use the wastes of agricultural production. Fruit body of mushrooms are appreciated, not only for texture and flavour but also for their chemical and nutritional properties (Manzi, Aguzzi, Vivanti, Paci, & Pizzoferrato, 1999). Mushrooms have also been reported as therapeutic foods, useful in preventing diseases such as hypertension, hypercholesterolemia, and cancer. These functional characteristics are mainly due to their chemical composition (Manzi, Aguzzi, & Pizzoferrato, 2001).

Heavy metal concentrations in mushroom are considerably higher than those in agricultural crop plants, vegetables and fruit. This suggests that mushrooms possess a very effective mechanism that enables them readily to take up some heavy metals from the ecosystem. The concentrations of trace elements in the fruiting bodies of fungi are primarily species-dependent. The concentrations were found to depend on the physiology of the species and particularly on its ecosystem pattern. It has proven rather difficult to determine the effects of

environmental factors on the heavy metals (Lepsova & Mejstrik, 1988).

In most countries, there is a well-established consumer acceptance of cultivated mushrooms (*Agaricus bisporus*, *Pleurotus* spp., *Lentinus edodes*, *Volvariella volvacea*, *Auricularia* spp., etc). However, wild edible mushrooms have been traditionally eaten only by specific groups of people and seasonally (Diez and Alvarez, 2001). Nevertheless, wild edible mushrooms are becoming increasingly important in our diet for their nutritional and pharmacological characteristics (Bobek et al., 1991; Breene, 1990; Crisan & Sands, 1978; Manzi et al., 2001). Therefore, it is necessary to investigate the levels of essential elements in wild mushrooms, because many wild edible mushroom species are known to accumulate high levels of several heavy metals, and mainly cadmium, mercury, copper and lead. Results from over one hundred original papers, dealing with heavy metals in edible mushrooms, show that cadmium, mercury and to a lesser extent lead are the metals of toxicological importance (Kalac and Svoboda, 2000).

In the Middle Black Sea region of Turkey, the climate is mild and rainy. Therefore, the seasons are normally wet with mild temperatures, especially; spring and autumn are suitable for fungal growth. People who live in this region of Turkey (Tokat) widely consume wild edible mushrooms because of their delicacy and abundance.

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Table 1
Operating conditions for mushroom samples in the microwave digestion system^a

Steps	Time (min)	Power (W)
1	2	250
2	2	0
3	6	250
4	5	400
5	8	550

^a Vent: 8 min.

Many studies have been carried out on the trace element contents of macrofungus in Turkey (Demirbaş, 2000; Tüzen, Özdemir, & Demirbaş, 1998; Yildiz, Karakaplan, & Aydın, 1998; Sesli and Tüzen, 1999; Sivrikaya, Bacak, Saracbaşı, Toroğlu, & Eroğlu, 2002). However, qualified studies have not been carried out in this area of Turkey.

The aim of this study is to determine Pb, Cd, Mn, Zn, Cu and Fe contents in the fruiting bodies of 10 wild mushroom species collected from Tokat, Turkey.

2. Materials and methods

The mushroom samples were collected from uncontaminated agricultural lands in Tokat, Turkey. The samples were dried at 105 °C for 24 h. Dried samples were homogenized using an agate homogenizer and stored in pre-cleaned polyethylene bottles until analysis.

All reagents were of analytical reagent grade unless otherwise stated. Double deionised water was used for all dilutions. HNO₃ and H₂O₂ were of suprapure quality (Merck).

For the elemental analysis, a Perkin-Elmer Analyst 700 atomic absorption spectrometer with deuterium background corrector was used in this study. Pb and Cd

in samples were determined by a HGA graphite furnace using Argon as inert gas. Other measurements were carried out in an air/acetylene flame.

For digestion, Milestone Ethos D microwave closed system was used in this study. Sample (0.25 g) was digested with 6 ml of HNO₃ (65%) and 1 ml of H₂O₂ (30%) in microwave digestion system for 23 min and final diluted to 25 ml with deionised water. A blank digest was carried out in the same way. Digestion conditions are given in Table 1.

3. Results and discussion

The habitat, edibility and the families of mushrooms used in this study are given in Table 2. The results of heavy metal concentrations in the mushroom species are shown in Table 3.

The heavy metal concentration in the mushrooms are mainly affected by acidic and organic matter content of their ecosystem and soil (Gast, Jensen, Bierling, & Haanstra, 1988). The uptake of metal ions in mushrooms is in many respects different from plants. For this reason the concentration variations of metals depend on mushroom species and their ecosystems (Seeger, 1982).

The highest iron content was 3904 mg/kg in *Fomes fomentarius* whereas the lowest iron content was 568 mg/kg in *Lepista nuda* (Fig. 1). Reported iron contents in mushrooms are much lower than ours except in three recent studies (Işiloğlu, Yılmaz, & Merdivan, 2001; Latiff, Daran, & Mohamed, 1996; Tüzen et al., 1998).

Copper content ranged from 18 to 54 mg/kg in the present study. The highest copper content was seen in *F. fomentarius* and the lowest in *Boletus appendiculatus* (Fig. 2). Copper contents in mushrooms higher than those in vegetables should be considered as a nutritional source of the element. Nevertheless, for people, bio-availability from mushrooms was reported to be low, due

Table 2
Families, habitat and edibility of mushroom species

No	Class, family and species of mushrooms	Habitat	Edibility
01	<i>Nectriaceae</i> <i>Nectria cinnabarina</i> (Tode.: Fr.) Fr.	On dead wood	Not edible
02	<i>Lycoperdaceae</i> <i>Calvatia utriformis</i> (Bull.: Pers) Jaap.	In pastures or on heaths, usually on sandy soil	Edible
03	<i>Bovista plumbea</i> Pers.: Per Pers.	Amongst short grass, on lawns	Edible
04	<i>Lycogola epidendron</i> Fr.	On trunks of coniferous	Poisonous
05	<i>Polyporaceae</i> <i>Coriolus versicolor</i> (L.: Fr.) Pilat.	On deciduous wood	Not edible
06	<i>Fomes fomentarius</i> (L.: Fr.) Fr.	On beech	Not edible
07	<i>Funallia trogii</i> (Berk.) Bend. et., Sing.	On dead twigs	Not edible
08	<i>Polyporus frondosus</i> (Mich.: Fr.)	On trunks of <i>Salix</i> spp.	Edible
09	<i>Boletaceae</i> <i>Boletus appendiculatus</i> Shaef.: Fr.	With broad-leaved trees	Edible
10	<i>Tricholomataceae</i> <i>Lepista nuda</i> (Bull.: Fr.) Cook.	In woodland hedgerows and garden	Edible

Table 3

Concentrations of Fe, Cu, Mn, Zn, Pb, and Cd of the mushroom samples analyzed (mg/kg, dry wt.) (mean \pm standard deviation), $n=5$

Mushroom samples	Fe	Cu	Mn	Zn	Pb	Cd
<i>Nectria cinnabarina</i>	1420 \pm 75	30 \pm 3	44 \pm 5	105 \pm 10	0.8 \pm 0.2	0.3 \pm 0.04
<i>Calvatia utriformis</i>	924 \pm 48	25 \pm 4	28 \pm 3	58 \pm 4	1.5 \pm 0.3	1.1 \pm 0.1
<i>Bovista plumbea</i>	2340 \pm 175	42 \pm 3	36 \pm 2	42 \pm 5	2.3 \pm 0.2	0.7 \pm 0.1
<i>Lycogola epidendron</i>	3562 \pm 282	21 \pm 2	41 \pm 3	71 \pm 6	1.2 \pm 0.1	1.4 \pm 0.1
<i>Coriolus lersicolor</i>	1905 \pm 108	47 \pm 3	28 \pm 2	39 \pm 3	0.9 \pm 0.1	0.5 \pm 0.07
<i>Fomes fomentarius</i>	3904 \pm 307	54 \pm 4	64 \pm 5	40 \pm 4	2.7 \pm 2.0	1.8 \pm 0.2
<i>Funalli trogii</i>	1665 \pm 190	32 \pm 4	33 \pm 2	25 \pm 2	1.6 \pm 0.2	1.4 \pm 0.1
<i>Polyporus frondosus</i>	2003 \pm 345	41 \pm 3	28 \pm 3	122 \pm 11	1.2 \pm 0.1	0.7 \pm 0.1
<i>Boletus appendiculatus</i>	1040 \pm 202	18 \pm 2	35 \pm 4	63 \pm 4	2.2 \pm 0.3	1.8 \pm 0.2
<i>Lepista nuda</i>	568 \pm 46	20 \pm 3	16 \pm 2	45 \pm 3	1.4 \pm 0.1	1.1 \pm 0.1

to limited absorption from the small intestine (Schellman, Hiltz, & Opitz, 1980). Copper levels in mushrooms are in agreement with those reported earlier (Demirbaş, 2000; Işiloğlu et al., 2001; Sesli & Tüzen, 1999; Tüzen et al., 1998).

In this study, the highest manganese content was 64 mg/kg, for the species *Fomes fomentarius*, whereas the lowest manganese content was 16 mg/kg, for the species *Lepista nuda* (Fig. 3). Manganese content is in good agreement with other studies (Demirbaş, 2001; Işiloğlu

et al., 2001; Jorhen & Sundström, 1995; Latiff & Daran, 1996; Sesli & Tüzen, 1999). In the previous works, the concentrations of manganese in mushrooms were large. But there are no great differences among the species.

Minimum and maximum values of zinc in the present study were 25 and 122 mg/kg. The highest and lowest levels were found in *Polyporus frondosus* and *Funalli trogii*, respectively (Fig. 4). Zinc is widespread among living organisms due to its biological significance.

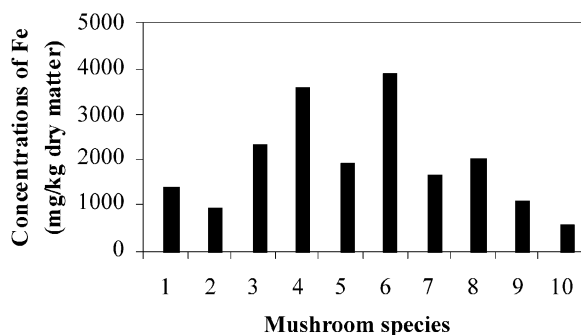


Fig. 1. Distribution of iron in mushroom species: 1, *Nectria cinnabarina*; 2, *Calvatia utriformis*; 3, *Bovista plumbea*; 4, *Lycogola epidendron*; 5, *Coriolus ersicolor*; 6, *Fomes fomentarius*; 7, *Funalli trogii*; 8, *Polyporus rondosus*; 9, *Boletus appendiculatus*; 10, *Lepista nuda*.

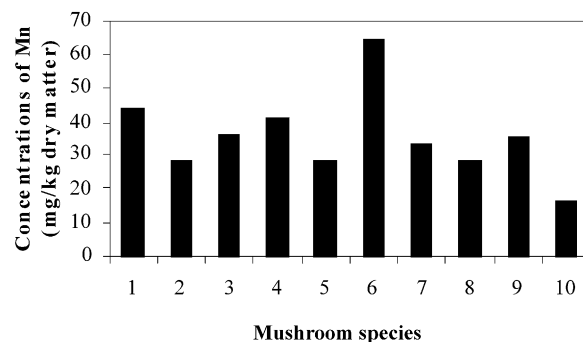


Fig. 3. Distribution of manganese in mushroom species: 1, *Nectria cinnabarina*; 2, *Calvatia utriformis*; 3, *Bovista plumbea*; 4, *Lycogola epidendron*; 5, *Coriolus ersicolor*; 6, *Fomes fomentarius*; 7, *Funalli trogii*; 8, *Polyporus rondosus*; 9, *Boletus appendiculatus*; 10, *Lepista nuda*.

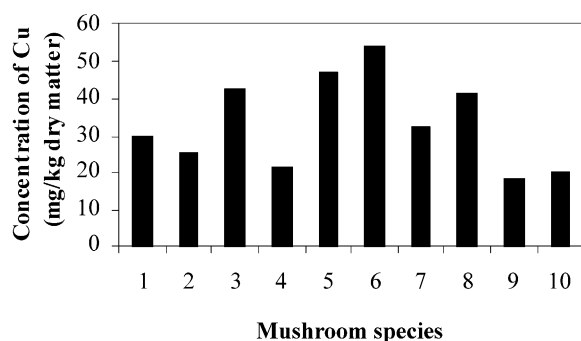


Fig. 2. Distribution of copper in mushroom species: 1, *Nectria cinnabarina*; 2, *Calvatia utriformis*; 3, *Bovista plumbea*; 4, *Lycogola epidendron*; 5, *Coriolus ersicolor*; 6, *Fomes fomentarius*; 7, *Funalli trogii*; 8, *Polyporus rondosus*; 9, *Boletus appendiculatus*; 10, *Lepista nuda*.

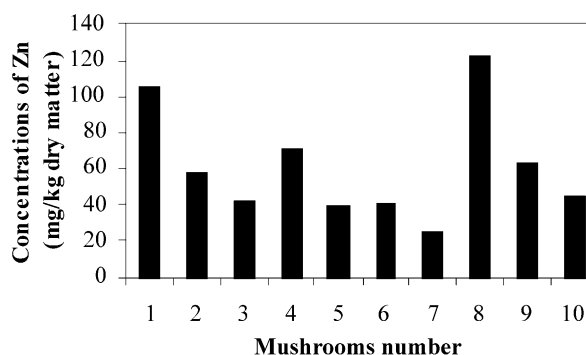


Fig. 4. Distribution of zinc in mushroom species: 1, *Nectria cinnabarina*; 2, *Calvatia utriformis*; 3, *Bovista plumbea*; 4, *Lycogola epidendron*; 5, *Coriolus ersicolor*; 6, *Fomes fomentarius*; 7, *Funalli trogii*; 8, *Polyporus rondosus*; 9, *Boletus appendiculatus*; 10, *Lepista nuda*.

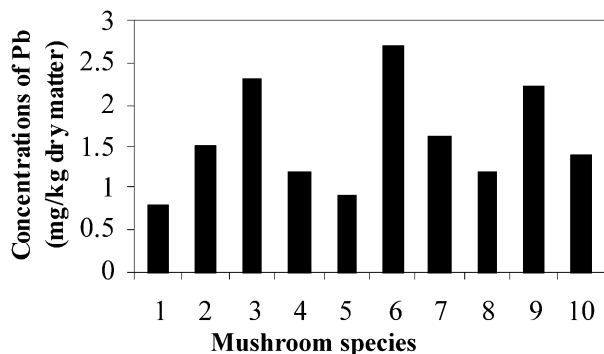


Fig. 5. Distribution of lead in mushroom species: 1, *Nectria cinnabarina*; 2, *Calvatia utriformis*; 3, *Bovista plumbea*; 4, *Lycogola epidendron*; 5, *Coriolus eriscolor*; 6, *Fomes fomentarius*; 7, *Funalli trogii*; 8, *Polyporus rondonus*; 9, *Boletus appendiculatus*; 10, *Lepista nuda*.

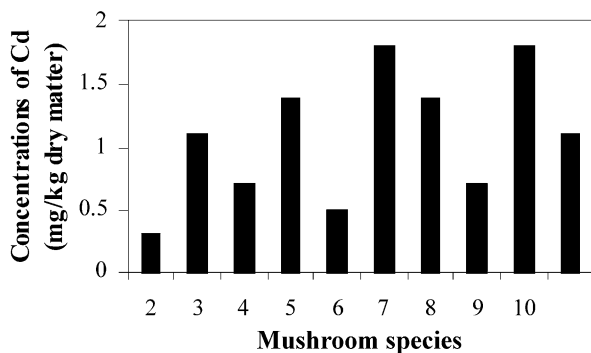


Fig. 6. Distribution of cadmium in mushroom species: 1, *Nectria cinnabarina*; 2, *Calvatia utriformis*; 3, *Bovista plumbea*; 4, *Lycogola epidendron*; 5, *Coriolus eriscolor*; 6, *Fomes fomentarius*; 7, *Funalli trogii*; 8, *Polyporus rondonus*; 9, *Boletus appendiculatus*; 10, *Lepista nuda*.

Content of zinc related in mushrooms ranges from 30 to 150 mg/kg (Kalac & Svoboda, 2000). Hence, zinc content in mushrooms of the present study is in agreement with the previous studies (Anderson, Lykke, Lange, & Bech, 1982; Kalac & Svoboda, 2000).

Lead concentrations in mushrooms of this study ranged from 0.8 to 2.7 mg/kg. The lowest mean levels were found in *Nectria cinnabarina* and the highest in *F. fomentarius* (Fig. 5). In the prior studies, data on lead concentrations in mushrooms are given in *Lycoperdon perlatum*, *Macrolpiota rhacodes*, and *L. nuda*, which are highly accumulating species. However, lead content in *L. nuda* was 1.4 mg/kg in this study.

The lowest cadmium content was 0.3 mg/kg in *N. cinnabarina* and the highest was 1.8 mg/kg in two species of *Boletus appendiculatus* and *F. fomentarius* (Fig. 6). Our data are in agreement with values given in the literature. Cadmium is accumulated mainly in kidneys and liver and its level in blood serum increases considerably following mushroom consumption.

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